

A Twist in the Tail: The Structural Basis of Kv7.2/Kv7.3 Channel Assembly through Helix D

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Epilepsy is one of the most prevalent neurological disorders, affecting around 65 million people worldwide¹. It is characterised by recurrent seizures, the severity of which can be influenced by genetic, environmental, or unknown factors. One in three individuals with epilepsy do not respond to current treatments², highlighting the need for new therapeutic targets and more effective therapies.

The Kv7 family of voltage-gated potassium channels (Kv7.1–Kv7.5), encoded by *KCNQ* genes (*KCNQ1*–*KCNQ5*), has been implicated in epilepsy³. Kv7.2 and Kv7.3 assemble to form a majority of the neuronal M-channel, which regulates neuronal excitability by generating the stabilising M-current that prevents excessive firing. Mutations in the *KCNQ* genes, particularly within the transmembrane and regulatory domains of Kv7.2 and Kv7.3, are strongly associated with epileptic phenotypes.

A novel mutation has been identified in Kv7.3 within the proposed assembly domain, Helix D, arising from a glutamine to leucine substitution (Q653L). This residue lies within the distal half of the C-terminal tail, a flexible region with limited structural data, and may reveal a new mechanism affecting channel assembly and function. While Helix D in Kv7.1 and Kv7.4 promotes tetramer formation^{4,5}, the corresponding region in Kv7.3 has been reported to favour monomeric and dimeric states⁴. Given Helix D in Kv7.3 has a lower sequence conservation within the Kv7 family, and its key role in assembly, further investigation is needed to clarify its contribution to M-channel formation.

Here, we present the first crystal structures of the Kv7.2 Helix D domain and a heteromeric Kv7.2/Kv7.3 Helix D complex, revealing key interfaces that likely mediate subunit association. Complementary biophysical analyses, including size-exclusion chromatography, multi-angle light scattering, mass photometry, and circular dichroism spectroscopy, show that Helix D forms a dynamic assembly capable of supporting distinct oligomeric states. These findings provide a structural framework for understanding Kv7 channel assembly and set the stage for ongoing cryo-EM studies of full-length Kv7.2/Kv7.3 channels.

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